

## Review Article

### Diagnostic Biomarkers in Cervical, Breast, and Prostate Cancers: A Review

Bernard kafor<sup>1</sup>, Charles Okolie<sup>2</sup> Gideon Ekpiri<sup>3</sup>

<sup>1</sup>Department of Medical Laboratory Science, Maduka University

<sup>2</sup>State University of Medical and Applied Sciences

<sup>3</sup>Studying at the. Anglia Ruskin University Cambridge

#### \*Corresponding author

Bernard kafor. Department of Medical Laboratory Science, Maduka University

Received: 02 February 2026

Accepted: 25 March 2026

Published: 22 April 2026

#### Copyright

© 2026 Bernard kafor

OPEN ACCESS

#### Abstract

Laboratory-based biomarkers are becoming more and more important in the diagnosis of cancer, supporting therapy selection, risk assessment, and early detection. Prostate, breast, and cervical cancers account for a significant amount of cancer-related morbidity and mortality worldwide, especially in Africa, where access to sophisticated testing and delayed diagnosis continue to be significant obstacles. With a focus on their applicability to laboratory medicine practice, this narrative review investigates both established and developing diagnostic biomarkers for these three cancers. Emerging liquid biopsy techniques like circulating tumor DNA (ctDNA), exosomal RNA, and circulating microRNAs are reviewed alongside validated tissue and molecular biomarkers, such as high-risk human papillomavirus (HPV) testing and p16/Ki-67 dual immunostaining for cervical cancer; oestrogen receptor (ER), progesterone receptor (PR), HER2, and multigene assays for breast cancer; and prostate-specific antigen (PSA) and urine-based molecular tests for prostate cancer. Important operational, analytical, and equity-related issues that impact implementation in environments with limited resources are highlighted. The research emphasizes how crucial diagnostic labs are to converting biomarker advancements into clinically and publicly significant cancer control plans throughout Africa.

**Keywords:** Diagnostic biomarkers; cervical cancer; breast cancer; prostate cancer; laboratory medicine; liquid biopsy; HPV; ER/PR; HER2; PSA

#### Introduction

Prostate, breast, and cervical cancers are the main causes of cancer-related death in Africa and are among the most frequently diagnosed cancers globally. Even while early detection and efficient screening have greatly decreased mortality in high-income settings, many African nations still struggle with late presentation, inadequate diagnostic capabilities, and unequal access to necessary laboratory services [1-4].

Modern oncological practice relies heavily on diagnostic biomarkers for screening, diagnosis, prognostication, and therapeutic decision-making.

These biomarkers include assays based on nucleic acids, multigene expression profiles, protein-based immunohistochemistry markers, and circulating tumor-derived analytes that can be found in blood or urine. It is crucial for laboratory professionals to comprehend the clinical value, performance characteristics, and implementation needs of these biomarkers [5-8].

This review summarizes the most recent research on diagnostic biomarkers for prostate, breast, and cervical cancers, with an emphasis on analytical validity, laboratory workflows, and applicability in African health systems.

**Table 1: Summary of Established and Emerging Biomarkers by Cancer Type**

Cancer	Established Biomarkers	Emerging Liquid Biopsy Biomarkers	Clinical Use	Limitations / Africa Context
Cervical	HPV DNA/mRNA, p16 <sup>INK4a</sup> /Ki 67	cfHPV DNA, circulating miRNAs	Screening, triage, recurrence monitoring	Limited follow-up systems; cost and lab infrastructure barriers
Breast	ER/PR, HER2	ctDNA, CTCs, exosomes	Subtyping, treatment selection, therapy monitoring	Limited access to molecular assays; liquid biopsy still research-stage
Prostate	PSA (total, velocity)	PCA3, ctDNA, CTCs	Screening, biopsy decision, prognosis	Low specificity of PSA; advanced molecular tests scarce in many regions

A comparative table with rows for each cancer (cervical, breast, prostate) and columns for biomarker categories.

(Adapted from the web)

## Methods

Diagnostic biomarkers pertinent to laboratory medicine in prostate, breast, and cervical malignancies were the subject of a narrative literature review. In addition to important oncology and pathology publications, worldwide recommendations, and current systematic reviews published up to 2025, electronic searches were conducted using PubMed and PubMed Central. Combinations of "diagnostic biomarkers," "molecular testing," "immunohistochemistry," "liquid biopsy," and "cancer type" were among the search phrases. Guidelines-endorsed biomarkers, research with obvious laboratory implications, and literature on implementation in low- and middle-income settings were prioritized.

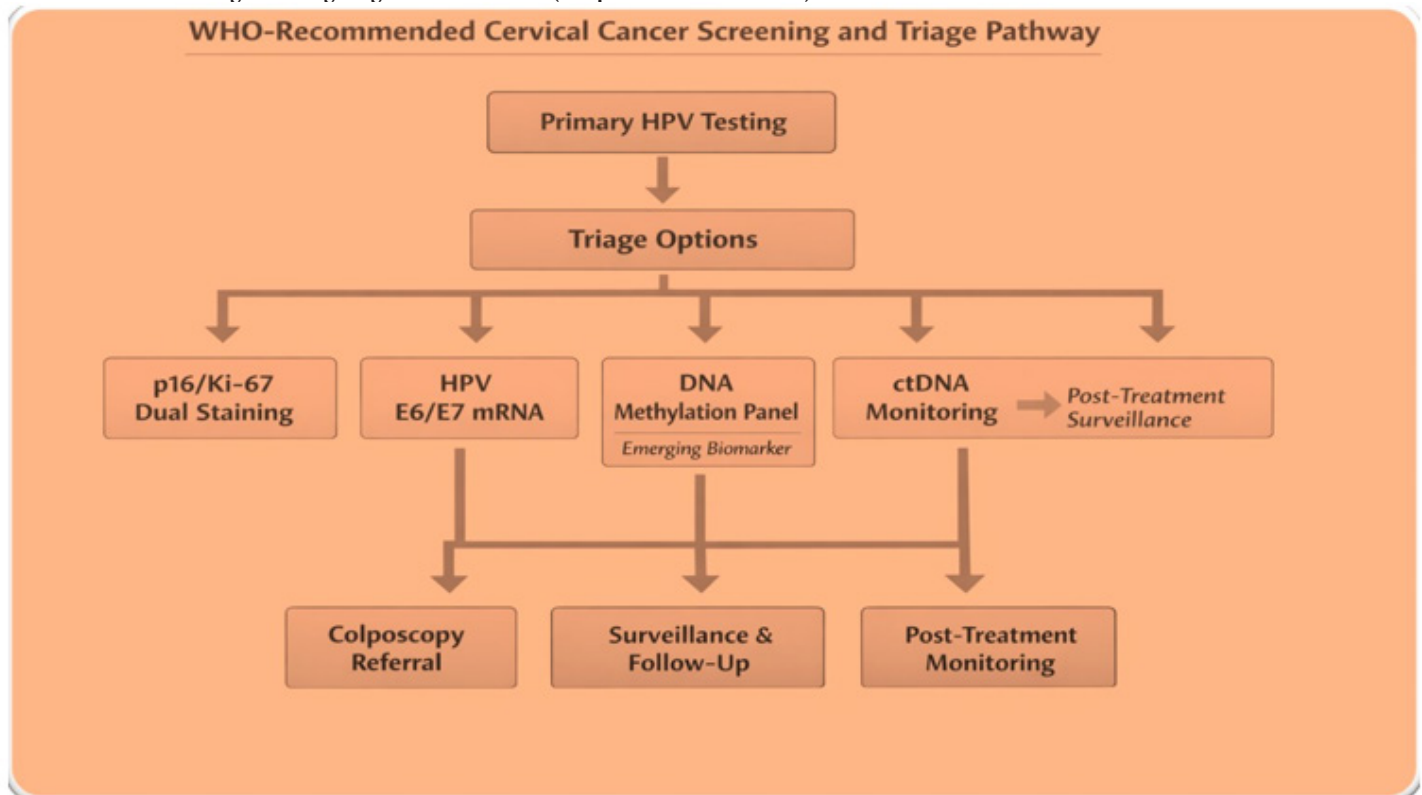
## Cervical Cancer Established laboratory biomarkers

The World Health Organization (WHO) recommends high-risk HPV testing as the preferred main screening approach, and it is the cornerstone of

modern cervical cancer screening. Cervical intraepithelial neoplasia grade 2 or worse (CIN2+) can be detected with great sensitivity using combined HPV DNA and mRNA assays, which surpass cytology alone. Longer screening intervals are made possible by primary HPV testing, which is compatible with high-throughput laboratory operations [9-12].

Dual immunostaining with p16<sup>INK4a</sup>/Ki-67 has become an established triage strategy for HPV-positive women, improving specificity for clinically significant cervical lesions by identifying HPV-induced cell cycle deregulation and active proliferation. This approach significantly reduces unnecessary colposcopy referrals, an important advantage in resource-limited health systems. HPV E6/E7 mRNA assays, by detecting transcriptionally active infections, offer higher specificity than HPV DNA testing and are increasingly incorporated into laboratory-based triage algorithms [19-20].

Cervical Cancer Screening and Triage Algorithm Flowchart (Adapted from the internet)



## Emerging biomarkers

The detection of circulating HPV DNA through liquid biopsy methods such as droplet digital PCR is being investigated as a tool for post-treatment surveillance and disease monitoring in invasive cervical cancer [21,22]. Furthermore, DNA methylation panels targeting viral and host gene sequences have shown potential as laboratory-based triage tools for diagnosing CIN2+ lesions [23-25]. Although many of these assays are still largely in the research stage, they may eventually serve as scalable alternatives to traditional cytology [26].

Conventional tumor markers, including squamous cell carcinoma antigen, are mostly utilized for disease surveillance and have limited diagnostic usefulness. Despite being outside of standard screening paradigms, immune biomarkers, such as PD-L1 expression, are becoming more and more important for directing treatment in advanced disease [27-32].

## Laboratory implementation considerations

In many African laboratories, issues with reagent availability, infrastruc-

ture limitations, quality assurance, and follow-up systems still exist despite the compelling evidence in favor of HPV-based screening [33-35]. Careful laboratory and public health coordination is necessary for the selection of suitable triage techniques and integration with immunization programs [36,37].

## Breast Cancer Core tissue biomarkers

The primary biomarkers for classifying breast cancer and choosing a course of treatment are still the estrogen receptor (ER), progesterone receptor (PR), and HER2. Decisions about endocrine therapy and HER2-targeted medications are made using immunohistochemistry (IHC) for ER/PR and IHC/ISH for HER2 [38]. In certain situations, chemotherapy decisions may be influenced by the Ki-67 proliferation index, which also provides information on prognosis [38]. Standardized testing and reporting are advised by guidelines [38,39].

## Genomic assays and multigene signatures

In ER-positive, HER2-negative early breast cancer, Oncotype DX, MammaPrint, Prosigna, EndoPredict, and other tools offer prognostic data and can forecast the value of adjuvant chemotherapy [38,39], these tools are increasingly used in guideline-directed decision-making. To prevent over-treatment, ASCO guidelines encourage the selective use of these assays [39].

## Liquid biopsy and circulating markers

For early detection, monitoring of minimal residual disease (MRD), and identification of actionable mutations (e.g., PIK3CA, ESR1) in metastatic illness, circulating tumor DNA (ctDNA) and circulating tumor cells (CTCs) are being intensively studied 40. In multi-cancer early detection investigations, ctDNA methylation panels have demonstrated a high sensitivity for cancer detection; they are currently being evaluated specifically for MRD detection and breast cancer screening 41. The metastatic scenario is where clinical adoption of response monitoring and mutation testing is expanding the fastest [40,41].

## Clinical application and limitations

Standardized Ki-67 scoring, interpretation of HER2-low status, analytic sensitivity of ctDNA for early disease, and cost/access difficulties for multigene tests continue to be challenges despite the validation and endorsement of ER/PR/HER2 testing and genomic assays [38,39,42]. An ongoing research objective is the incorporation of liquid biomarkers into routine early detection [40-42].

## Prostate Cancer PSA and derivatives

The most popular biomarker for identifying and tracking prostate cancer remains prostate specific antigen (PSA). PSA testing in many screening

programs has contributed to earlier detection and lower prostate cancer mortality; however, it has well established limitations, including low specificity, risk of overdiagnosis, and risks related to biopsy procedures. PSA derivatives such as PSA velocity, PSA density, and percentage of free/total PSA, along with multivariable risk calculators, offer modest improvements in discriminatory performance compared with PSA alone [42].

## Urine and tissue molecular tests

Urinary molecular tests, including urinary mRNA panels such as MyProstateScore (MiPS), SelectMDx, and ExoDx Prostate Intelliscore, use combinations of biomarkers (e.g., TMPRSS2 ERG fusion transcripts and PCA3) to improve specificity for clinically significant disease and help guide decisions about repeat biopsies. Tissue based genetic assay panels such as Prolaris, Decipher, and Oncotype DX Prostate provide prognostic information that informs treatment decision making between definitive therapy and active surveillance [42-45].

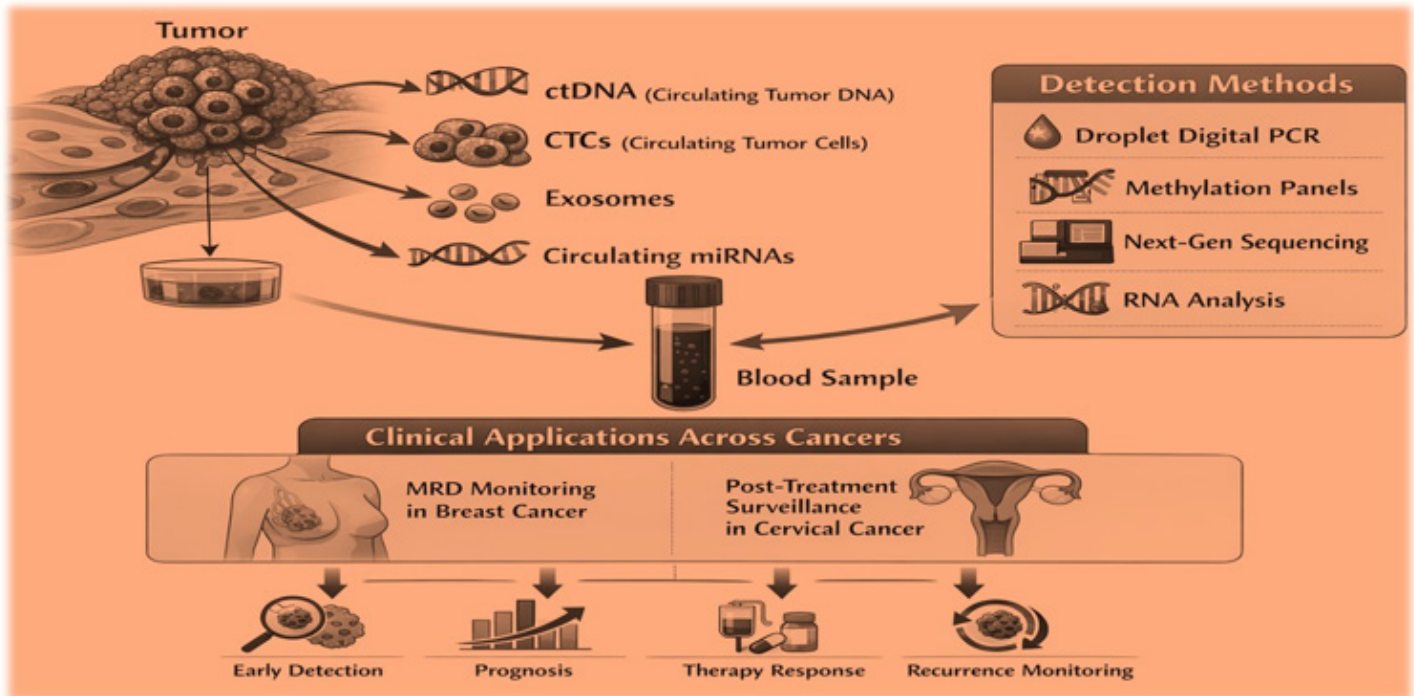
## Composite and commercial risk scores

Serum based composite scores such as the Prostate Health Index (PHI) and the 4Kscore combine PSA and kallikrein markers (including total PSA, free PSA, intact PSA, and human kallikrein 2) to stratify risk and reduce unnecessary biopsies by improving differentiation between benign disease and clinically significant prostate cancer. These tests have been validated in multiple cohorts and are increasingly available in clinical practice [42-44].

## Liquid biopsy and emerging biomarkers

Liquid biopsy techniques, including analysis of circulating tumor DNA (ctDNA) methylation, extracellular vesicle RNA, and microRNA (miRNA) signatures in urine and blood, are emerging as noninvasive methods to detect clinically important disease and monitor disease course. While promising, many candidates remain under clinical validation, and integration of.

### Components of Liquid Biopsy Techniques



- A schematic diagram illustrating key elements (e.g., ctDNA, CTCs, exosomes, circulating miRNAs) with icons or labeled arrows showing their origins from tumors, detection methods (e.g., droplet digital PCR, methylation panels), and applications across cancers (e.g., MRD monitoring in breast, post-treatment surveillance in cervical).

## Analytical and clinical validity

The analytical variability of assays (preanalytical variables, assay platforms, thresholds) is a major obstacle to translation [50]. Cost-effectiveness evaluations, prospective clinical validation, and thorough analytical validation are necessary for high-impact adoption [51,52]. There is an urgent need for harmonized reporting, assay performance measures, and standardized sample management [52].

## Equity, access, and guideline alignment

The availability of high-value biomarkers (HPV testing, ER/PR/HER2, PSA) varies by area [53]. In order to lessen global inequities, WHO guidelines recommend scalable cervical screening programs (HPV primary testing when possible, alternatives where necessary) [54]. The equitable application of modern biomarkers and genetic testing is influenced by factors such as cost, laboratory infrastructure, and physician training [55].

## Future Directions and Research Priorities

1. Extensive prospective cohort studies to verify multi-analyte liquid tests and ctDNA methylation for early detection and MRD in these malignancies.
2. Standardization programs (international consortia) for test preanalytics and reporting
3. Research on the cost-effectiveness and execution of HPV testing scale-up and p16/Ki-67 triage techniques in low- and middle-income countries.
4. Combining AI with multi-omics (genomics, methylomics, proteomics) to increase diagnostic precision while maintaining clinical interpretability and regulatory supervision.

## Conclusions

Prostate, breast, and cervical cancer detection and treatment have been revolutionized by diagnostic biomarkers. Current clinical practice and guidelines are supported by established tissue and molecular indicators (HPV testing and p16/Ki-67 for cervical cancer; ER/PR/HER2 and multi-gene assays for breast cancer; PSA and composite urine/blood panels for prostate cancer). The quick development of liquid biopsy technologies, especially ctDNA and methylation panels, has great potential for early detection and customized monitoring; however, before widespread routine adoption, issues with analytic standardization, prospective validation, cost-effectiveness, and equitable access must be resolved. To turn promising biomarkers into population benefits, more cooperative research and practical implementation studies will be necessary.

## References

1. Bray F, Parkin DM. (2022) Cancer in sub-Saharan Africa in 2020: a review of current estimates of the national burden, data gaps, and future needs. *Lancet Oncol.* 23: 719–728. doi:10.1016/S1470-2045(22)00270-4.
2. World Health Organization Regional Office for Africa. (2025) Cancer: health topics. Brazzaville: WHO Regional Office for Africa. Available from: <https://www.afro.who.int/health-topics/cancer>
3. Sharma R, Aashima, Nanda M, Fronterre C, Sewagudde P, Ssentongo AE, et al. (2022) Mapping cancer in Africa: a comprehensive and comparable characterization of 34 cancer types using estimates from GLOBOCAN 2020. *Front Public Health.* 10: 839835. doi:10.3389/fpubh.2022.839835.
4. Kibret AA, Jiang H, Yang H, Liu C. (2024) Patient journey and timeliness of care for patients with breast cancer in Africa: a scoping review protocol. *BMJ Open.* 14: e081256. doi:10.1136/bmjopen-2023-081256.
5. Simon RM, Paik S, Hayes DF. (2009) Use of archived specimens in evaluation of prognostic and predictive biomarkers. *J Natl Cancer Inst.* 101: 1446–1452. doi:10.1093/jnci/djp335.
6. Cree IA, Deans Z, Ligtenberg MJL, Normanno N, Edsjö A, Rouleau

- E, et al. (2014) Guidance for laboratories performing molecular pathology for cancer patients. *J Clin Pathol.* 67: 923–931. doi:10.1136/jclinpath-2014-202404.
7. Diamandis EP. (2012) The failure of protein cancer biomarkers to reach the clinic: why, and what can be done to address the problem? *BMC Med.* 10: 87. doi:10.1186/1741-7015-10-87.
8. Ignatiadis M, Sledge GW, Jeffrey SS. (2021) Liquid biopsy enters the clinic—implementation issues and future challenges. *Nat Rev Clin Oncol.* 18: 297–312. doi:10.1038/s41571-020-00457-x.
9. World Health Organization. (2021) WHO guideline for screening and treatment of cervical pre-cancer lesions for cervical cancer prevention. 2nd ed. Geneva: World Health Organization.
10. World Health Organization. (2020) Global strategy to accelerate the elimination of cervical cancer as a public health problem. Geneva: World Health Organization.
11. Ronco G, Dillner J, Elfström KM, Tunesi S, Snijders PJF, Arbyn M, et al. (2014) Efficacy of HPV-based screening for prevention of invasive cervical cancer: follow-up of four European randomised controlled trials. *Lancet.* 383: 524–532.
12. Arbyn M, Snijders PJF, Meijer CJLM, Berkhof J, Cuschieri K, Kocjan BJ. (2015) Which high-risk HPV assays fulfil criteria for use in primary cervical cancer screening? *Clin Microbiol Infect.* 21: 817–826.
13. Schmidt D, Bergeron C, Denton KJ, Ridder R. (2011) European CINtec Cytology Study Group. p16/Ki-67 dual-stain cytology in the triage of ASC-US and LSIL Papanicolaou cytology. *Cancer Cytopathol.* 119: 158–166.
14. Wentzensen N, Fetterman B, Castle PE, Schiffman M, Wood SN, Stiemerling E, et al. (2015) p16/Ki-67 dual stain cytology for detection of cervical precancer in HPV-positive women. *J Natl Cancer Inst.* 107: djv257.
15. Ikenberg H, Bergeron C, Schmidt D, Griesser H, Alameda F, Angeloni C, et al. (2013) Screening for cervical cancer precursors with p16/Ki-67 dual-stained cytology: results of the PALMS study. *J Natl Cancer Inst.* 105: 1550–1557.
16. Dijkstra MG, Heideman DAM, van Kemenade FJ, Hogewoning CJ, Hesselink AT, Verkuijten MCM, et al. (2010) p16INK4a immunocytochemistry in liquid-based cytology: a triage tool for HPV-positive women. *Br J Cancer.* 102: 936–943.
17. Kuhn L, Denny L, Pollack A, Lorincz A, Richart RM, Wright TC Jr. (2000) Human papillomavirus DNA testing for cervical cancer screening in low-resource settings. *J Natl Cancer Inst.* 92: 818–825.
18. Ratnam S, Coutlée F, Fontaine D, Bentley J, Escott N, Ghatage P, et al. (2011) Clinical performance of the APTIMA HPV mRNA assay compared with Hybrid Capture 2 HPV DNA test. *J Clin Microbiol.* 49: 557–563.
19. Arbyn M, Snijders PJ, Meijer CJ, Berkhof J, Cuschieri K, Kocjan BJ. (2015) Which high-risk HPV assays fulfil criteria for use in primary cervical cancer screening? *Clin Microbiol Infect.* 21: 817–826.
20. Wentzensen N, Clarke MA, Perkins RB. (2020) Impact of HPV mRNA testing on cervical cancer screening and triage. *Lancet Oncol.* 21: e278–e287.
21. Jensen SB, Thangarajah F, Jeppesen MM, et al. (2024) Circulating HPV DNA as a liquid biopsy biomarker in cervical cancer: potentials and challenges. *Int J Gynecol Cancer.* 34: 307–316. doi:10.1136/ijgc-2023-004873.
22. Chan KYK, Tsoi H, Shi J, et al. (2019) Liquid biopsy of HPV DNA in plasma from cervical cancer patients using droplet digital PCR. *J Clin Virol.* 113: 11–17. doi:10.1016/j.jcv.2019.02.013.
23. Dijkstra MG, van der Ploeg CPB, Steenbergen RDM, et al. (2019) DNA methylation markers for detection of high-grade cervical intraepithelial neoplasia and cervical cancer. *Br J Cancer.* 120: 363–375. doi:10.1038/s41416-019-0593-4.
24. Salta S, Stoler MH, Monsonego J, et al. (2023) The use of HPV DNA methylation tests in cervical screening: clinical performance and util-

- ity. *Clin Epigenetics*. 15: 115. doi:10.1186/s13148-023-01537-2.
25. Zhang S, Tan X, Yang F, et al. (2022) Host cell gene and HPV DNA methylation markers: a promising triage approach for cervical cancer screening. *Front Oncol*. 12: 843547.
  26. Jerónimo C, Castle PE. (2025) Integrating molecular triage strategies to optimize cervical cancer screening programs. *Life (Basel)*. 15: 367.
  27. Salazar-Ortega S, García-Ramírez J, Ruiz-García E, et al. (2017) Squamous cell carcinoma antigen as a tumor marker in cervical cancer: clinical utility and limitations. *Int J Gynecol Cancer*. 27: 1245–1252. doi:10.1097/IGC.0000000000001013.
  28. Bignotti E, Ravaggi A, Signorelli M, et al. (2012) Squamous cell carcinoma antigen in cervical cancer: predictive value for response and recurrence. *Gynecol Oncol*. 125: 330–334. doi:10.1016/j.ygyno.2012.01.025.
  29. Deng L, Li J, Wang J, et al. (2018) Clinical value of squamous cell carcinoma antigen in monitoring treatment response and recurrence of cervical squamous cell carcinoma. *Oncol Lett*. 15: 9933–9938. doi:10.3892/ol.2018.8563.
  30. Chung HC, Ros W, Delord JP, et al. (2019) Efficacy and safety of pembrolizumab in previously treated advanced cervical cancer: results from the KEYNOTE-158 study. *J Clin Oncol*. 37: 1470–1478. doi:10.1200/JCO.18.01265.
  31. Burke KP, Ferris RL. (2017) Immune checkpoint inhibition in head and neck squamous cell carcinoma: clinical relevance of PD-L1 expression. *Oral Oncol*. 68: 20–27. doi:10.1016/j.oraloncology.2017.03.002.
  32. Cohen AC, Roane BM, Leath CA. (2020) Novel therapeutics for recurrent cervical cancer: moving towards personalized immunotherapy. *Gynecol Oncol*. 156: 473–482. doi:10.1016/j.ygyno.2019.12.003.
  33. Egbon M, Ojo T, Aliyu A, et al. (2023) Challenges faced in implementing HPV testing for cervical cancer screening across public health programmes in sub-Saharan Africa: constraints related to resources, infrastructure and follow-up systems. *BMJ Open*. 13: e065074. doi:10.1136/bmjopen-2022-065074.
  34. Fitzpatrick C, Tota J, Chevarie Davis M, et al. (2020) Opportunities and challenges for introducing HPV testing in sub-Saharan Africa: laboratory infrastructure, supply availability, and quality issues. *BMC Public Health*. 20: 60.
  35. Ginsburg O, Bray F, Coleman MP, et al. (2011) Challenges of effective cervical cancer screening and patient follow-up in low-income countries: resource and system-level constraints. *Int J Equity Health*. 10: 27.
  36. Smith MA, Mbatha JW, Ndlela B, et al. (2023) Integrating HPV vaccine delivery with adolescent health services: experiences, enablers, and barriers from African countries. *Vaccine*. 42: S45–S48.
  37. Amponsah-Dacosta E, Kagina BM, Olivier J. (2020) Health systems constraints and facilitators of HPV immunization programmes in sub-Saharan Africa: a systematic review. *Health Policy Plan*. 35: 701–717. doi:10.1093/heapol/czaa017.
  38. Wolff AC, Hammond MEH, Allison KH, et al. (2018) Human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists guideline focused update. *J Clin Oncol*. 36: 2105–2132.
  39. Andre F, Ciruelos E, Juric D, et al. (2022) Biomarkers for adjuvant endocrine and chemotherapy in early-stage breast cancer: ASCO guideline update. *J Clin Oncol*. 40: 1816–1837.
  40. Zeng C, Zhang J. (2022) A narrative review of five multigenetic assays in breast cancer. *Transl Cancer Res*. 11: 897–907. doi:10.21037/tcr-21-1920.
  41. Klein EA, Richards D, Cohn A, et al. (2025) ctDNA in early cancer detection: promise and limitations. *Eur J Med Res*. 30: 30.
  42. Schiavone ML, Scarpitta R, Ravera F, Bleve S, Reduzzi C, Di Cocco F, et al. (2025) Liquid biopsy in breast cancer: redefining precision medicine. *J Liquid Biopsy*. 9: 100312. doi:10.1016/j.jlb.2025.100312.
  43. Loeb S, Catalona WJ. (2013) Use of prostate-specific antigen testing for screening and early diagnosis of prostate cancer. *J Urol*. 189: S1–S7.
  44. Pepe P, Garufi A. (2017) Prognostic biomarkers used for localized prostate cancer management: a systematic review. *Prog Urol*. 27: 398–411.
  45. Paller CJ, Antonarakis ES. (2025) Biomarkers in localized prostate cancer: from diagnosis to treatment. *Int J Mol Sci*. 26: 7667.
  46. Alix-Panabières C, Pantel K. (2021) Liquid biopsy: from discovery to clinical application. *Cancer Discov*. 11: 858–873.
  47. Bidard FC, Michiels S, Riethdorf S, et al. (2018) Circulating tumor cells in breast cancer patients treated by neoadjuvant chemotherapy: a meta-analysis. *J Natl Cancer Inst*. 110: 560–567.
  48. Ignatiadis M, Sledge GW, Jeffrey SS. (2021) Liquid biopsy enters the clinic: implementation issues and future challenges. *Nat Rev Clin Oncol*. 18: 297–312.
  49. Liu MC, Oxnard GR, Klein EA, et al. (2020) Sensitive and specific multi-cancer detection and localization using methylation signatures in cell-free DNA. *Ann Oncol*. 31: 745–759.
  50. Lennon AM, Buchanan AH, Rego SP, et al. (2020) Path to clinical implementation of cell-free DNA-based detection of multiple cancers. *J Clin Oncol*. 38: e15001.
  51. Merker JD, Oxnard GR, Compton C, et al. (2018) Circulating tumor DNA analysis in patients with cancer: American Society of Clinical Oncology and College of American Pathologists joint review. *J Clin Oncol*. 36: 1631–1641.
  52. Pennell NA, Arcila ME, Gandara DR, West H. (2019) Biomarker testing for patients with advanced non-small cell lung cancer: real-world issues and tough choices. *Am Soc Clin Oncol Educ Book*. 39: 531–542.
  53. Rolfo C, Mack P, Scagliotti GV, et al. (2021) Liquid biopsy for advanced non-small cell lung cancer: a consensus statement from the International Association for the Study of Lung Cancer. *J Thorac Oncol*. 16: 1647–1662.
  54. Lampignano R, Neumann MHD, Weber S, et al. (2020) Multicenter evaluation of circulating cell-free DNA extraction and downstream analyses for the development of standardized (pre)analytical workflows. *Clin Chem*. 66: 149–160.
  55. Sung H, Ferlay J, Siegel RL, et al. (2021) Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*. 71: 209–249.
  56. World Health Organization. (2021) WHO guideline for screening and treatment of cervical pre-cancer lesions for cervical cancer prevention. 2nd ed. Geneva: WHO.
  57. Brisson M, Kim JJ, Canfell K, et al. (2020) Impact of HPV vaccination and cervical screening on cervical cancer elimination: a comparative modelling analysis in 78 low-income and lower-middle-income countries. *Lancet*. 395: 575–590.

**Cite this article:** \*Bernard kafor and Charles Okolie. (2026) Diagnostic Biomarkers in Cervical, Breast, and Prostate Cancers: A Review. *Advance Medical & Clinical Research*. 7 (1): 315-319.

**Copyright:** ©2026 Bernard kafor. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.