

Research Article

Detection of viral and bacterial infections by means of Filmarray at a Tertiary Hospital in GreeceMourtzikou A^{1*}, Seitopoulou C^{1,2}, Stamouli M³, Kalliora G⁴, Kimouli M⁵¹Laboratory of Molecular Diagnostics, GHNP "Agios Panteleimon", Piraeus, Greece²Laboratory of Biopathology, Nikea Primary Healthcare Center, 2nd YPE, Nikea, Piraeus, Greece³Biochemistry Laboratory, Naval and Veterans Hospital of Athens, Athens, Greece⁴Faculty of Biology, National and Kapodistrian University of Athens (EKPA), Athens, Greece⁵Microbiology Laboratory, GHNP "Agios Panteleimon", Piraeus, Greece***Corresponding author**

Mourtzikou A. Laboratory of Molecular Diagnostics, GHNP "Agios Panteleimon", Piraeus, Greece

Received: 15 February 2026**Accepted:** 20 February 2026**Published:** 13 March 2026**Copyright**

© 2026 Mourtzikou A

OPEN ACCESS

Abstract**Introduction:** Respiratory Viruses (RVs) and bacteria are the main cause of pulmonary infections that affect the community population. They consist major public health concern with high impact in the health care systems. Their seasonal circulation is of great importance for the public health sector.**Aim:** The aim of the present study is to investigate the presence of RVs and bacteria in patients who visited the outpatients department, as well as in patients hospitalized at the GHNP "Agios Panteleimon" during 2024.**Materials and Methods:** In the study 483 patients, 272 male (56.3%) and 271 female (43.7%) were included. Patient age ranged from 1 to 98 years (mean 69.46 y; SD 13.49 y). Samples were collected from January to December 2024. Identification of bacteria (*Bordetella pertussis*, *Mycoplasma pneumoniae*), and RVs (*Adenovirus*, *Coronavirus 229E*, *Coronavirus HKU 1*, *Coronavirus NL 63*, *Coronavirus OCH3*, *Coronavirus OCK3*, *Human metapneumovirus*, *Influenza A*, *Influenza AH3*, *Influenza B*, *Parainfluenza 3 virus*, *Parainfluenza 1 virus*, *RSV* and *SARS-CoV-2*) was performed by the FILMARRAY Multiplex PCR System.**Results:** From epidemiological aspect maximum peak occurred in March, following Christmas and winter vacation, as well as the effect of low temperature and elevated humidity levels during winter months. During spring and early summer we observed also a significant increase in infections, due to climate change in the country. Moreover, a significant elevation of infections was observed during August, September and October, following summer vacation, as well as the return to school and work, and the consequent close contact in classrooms and working areas. Statistical significance between the result positivity with patient gender or patient age was not observed.**Conclusions:** Our study revealed seasonality of the infections. Early detection and surveillance of viral and bacterial infection can help in better organization, management and economic efficiency for the health care system.**Keywords:** Filmarray, Multiplex PCR, viruses, bacteria, influenza, Corona virus, RSV**Introduction**

Rapid and accurate identification of viral and bacterial pathogens in clinical specimens remains a central challenge in contemporary infectious disease diagnostics [1-3]. Traditional culture-based and immunoassay methods are often limited by long turnaround times, variable sensitivity, and dependence on specialist laboratory infrastructure, which can delay appropriate clinical management and antimicrobial stewardship decisions [4-7]. Molecular multiplex assays- also known as syndromic diagnostic tests- have emerged as a transformative approach in this context, enabling simultaneous detection of multiple targets with high analytical sensitivity and specificity directly from patient samples [8-15].

The FilmArray system (BioFire Diagnostics, bioMérieux) is a fully automated multiplex polymerase chain reaction (PCR) platform that integrates nucleic acid extraction, amplification, and detection within a closed pouch format. This design significantly reduces hands-on time and contamination risk while providing results in approximately one hour, facilitating syndromic pathogen identification across a range of clinical syndromes [16-20]. The FilmArray Respiratory Panel (RP and RP2.1) can detect an extensive panel of viral and bacterial respiratory pathogens, demonstrating robust performance in comparative studies and enabling identification of co-infections that might be missed by conventional methods (e.g., *adenoviruses*, *influenza viruses*, *common coronaviruses*, *Mycoplasma pneumoniae*, *Bordetella pertussis*) and support timely epidemiological insights [21-24]. The rapid turnaround and broad target range have also been eval-

uated in disease-specific contexts such as meningitis/encephalitis, where FilmArray's multiplex PCR panel for cerebrospinal fluid improved pathogen detection rates compared with standard diagnostics, although challenges remain in interpretation of false positives and negatives [25, 26].

In bloodstream infections, the Film Array Blood Culture Identification 2 (BCID2) panel has been shown to identify bacterial and fungal pathogens along with key antimicrobial resistance markers with high concordance to conventional methods, while substantially reducing time to result, which is critical for optimizing antimicrobial therapy [3, 27, 28]. Multiple studies in respiratory and bloodstream infection settings have confirmed that implementation of Film Array panels can influence clinical decision making, reduce time to appropriate therapy, and enhance diagnostic yield compared with traditional culture-based methods [29, 30].

Given the continued evolution of multiplex molecular diagnostics and their increasing integration into routine hospital workflows, it is essential to characterize the performance, clinical utility, and limitations of Film Array technology in the detection of viral and bacterial infections within specific healthcare settings. In this retrospective study, we aimed to characterize the spectrum of respiratory viral and bacterial pathogens identified by the Film Array system in a Greek tertiary hospital during 2024 and to highlight the potential role of syndromic testing in hospital-based surveillance.

Objective

The aim of the present retrospective study is to investigate the presence of RVs and bacteria in patients who visited the outpatients department, as well as in patients hospitalized at a tertiary hospital in Piraeus region, Greece, during 2024.

Material and methods

In the present retrospective study 483 patients, 272 male (56.3%) and 211 female (43.7%) were included. Patient age ranged from 1 to 98 years (mean 69.46 y; SD 13.49 y). Samples were collected from January to December as follows: January 18 samples, February 54, March 75, April 53, May 53, June 50, July 26, August 41, September 37, October 42, November 26 and December 8 samples. Among them 3 referrals were from the Cardiology department, 4 from the Neonatology, 353 from the Pulmonology, 3 from the Neurology, 105 from the Pathology, 3 from the Pediatrics, 1 from the Psychiatry, 1 from the Surgery, 1 from the ICU, 3 from the ED and 5 outpatients. The identification of bacteria and viruses was performed by means of FILM ARRAY Multiplex PCR System.

The study was conducted in accordance with the Declaration of Helsinki. All data were collected and processed in compliance with the General Data Protection Regulation (GDPR). Participant anonymity and confidentiality were strictly maintained, and no personally identifiable information was recorded. Informed consent was not required due to the retrospective nature of the study.

Results

Out of 483 patients, 272 were male (56.3%) and were 211 female (43.7%), Figure 1.

Single infections involving one pathogen were detected in 86 patients (Table 1), while mixed infections involving two or more pathogens were detected in 44 patients (Table 2). The annual distribution of the samples included in the study is presented in Table 3.

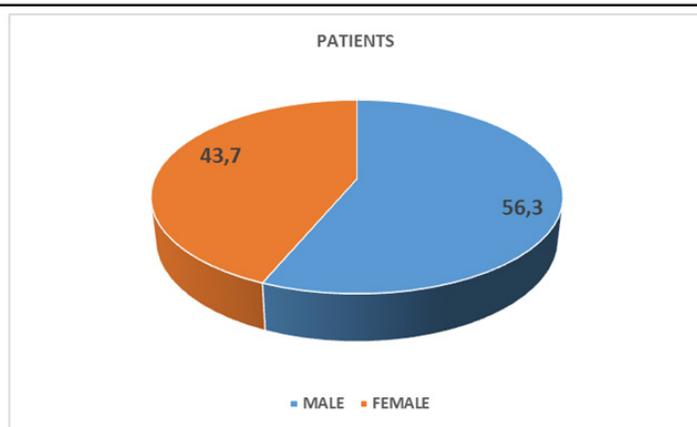


Figure 1. Gender distribution of the study population.

Table 1: Number of patients samples with single infections, involving only one pathogen.

SINGLE INFECTIONS	
Adenovirus	4
Bordetella pertussis	3
Corona virus 229E	2
Corona virus HKU1	1
Corona virus OCH3	1
Corona virus OCK3	2
Human metapneumovirus	11
Influenza A,	9
Influenza AH3	1
Influenza B	4
Parainfluenza 3 virus	9
Parainfluenza 1 virus	3
Mycoplasma pneumoniae	2
RSV	8
SARS-CoV-2	17
Infulenza A H1-2002	1
Corona virus NL 63	5
Corona virus OC43	3
TOTAL	86

Table 2: Number of patient samples with mixed infections, involving two or more pathogens.

MIXED INFECTIONS	
Influenza B/Bordetella pertussis	1
Adenovirus/Corona virus	1
Adenovirus/Corona virus 229 E	1
Corona virus OC43/Corona virus_NL_63	1
Rhinovirus/Enterovirus	31

Adenovirus/Rhinovirus/Enterovirus	1
Corona virus OC43/Rhinovirus/Enterovirus	6
Corona virus 229E/Rhinovirus/Enterovirus	1
Rhinovirus/Enterovirus/Parainfluenza 1	1
TOTAL	44

Table 3: Annual distribution of the samples included in the study.

NUMBER OF SAMPLES	
JANUARY	18
FEBRUARY	54
MARCH	75
APRIL	53
MAY	53
JUNE	50
JULY	26
AUGUST	41
SEPTEMBER	37
OCTOBER	42
NOVEMBER	26
DECEMBER	8
TOTAL	483

353 (73.1 %) samples were negative and 130 (26.9 %) were positive as follows: 4 Adenovirus cases, 3 Bordetella pertussis, 2 Corona virus 229E, 1 Corona virus HKU1, 1 Corona virus OCH3, 2 Corona virus OCK3, 11 Human metapneumovirus, 9 Influenza A, 1 Influenza AH3, 4 Influenza B, 9 Parainfluenza 3 virus, 3 Parainfluenza 1 virus, 2 Mycoplasma pneumoniae, 8 RSV, 17 SARS-CoV-2, 1 Influenza A H1-2002, 5 Corona virus NL 63 and 3 Corona virus OC43 cases.

Among the positive samples, 44 cases of mixed infection were detected as follows: 1 Influenza B/Bordetella pertussis, 1 Adenovirus/Corona virus, 1 Adenovirus/Corona virus 229 E, 1 Corona virus OC43/Corona virus NL-63, 31 Rhinovirus/Enterovirus, 1 Adenovirus/Rhinovirus/Enterovirus, 6 Corona virus OC43/Rhinovirus/Enterovirus, 1 Corona virus 229E/Rhinovirus/Enterovirus and 1 Rhinovirus/ Enterovirus/ Parainfluenza 1 case.

We did not detect any statistical significance between the result positivity with patient gender or patient age.

Discussion

Our study revealed seasonality of the infections. Maximum peak occurred in March, following Christmas and winter vacation, as well as the effect of low temperature and elevated humidity levels during winter months. During spring and early summer (April, May and June) we observe also a significant increase of the infections, due to climate change. Moreover, a significant elevation is observed during August, September and October, following summer vacation, as well as the return to school and work, and the consequent close contact in classrooms and working areas.

The present retrospective study provides important real-world evidence on the epidemiology of respiratory viral and atypical bacterial infections detected using the Film Array multiplex PCR platform in a Greek tertiary

hospital during 2024. By characterizing pathogen distribution, co-infections, departmental burden, and seasonal trends, our findings contribute to the growing international literature supporting the role of syndromic molecular diagnostics in contemporary infectious disease practice.

Overall, approximately one quarter of analyzed respiratory specimens yielded a positive result, while the majority remained negative. This diagnostic yield is in line with international hospital-based evaluations of syndromic multiplex respiratory panels, which commonly report positivity rates between 20% and 40% depending on patient selection and seasonal circulation patterns [16, 21].

The high proportion of negative samples may reflect the multi factorial nature of respiratory syndromes in hospitalized elderly populations, the timing of sampling in relation to symptom onset, and the restricted target spectrum of the assay, underscoring the importance of integrating molecular results with clinical assessment and diagnostic stewardship principles [19].

A notable characteristic of our population was the advanced mean age (≈ 69 years) and the predominance of referrals from Pulmonology and Internal Medicine wards, highlighting the substantial burden of respiratory viral infections among older hospitalized adults. This finding aligns with international evidence showing that common respiratory viruses, including influenza, RSV, and seasonal corona viruses, remain major drivers of hospitalization and healthcare strain in elderly patients beyond the pandemic period [21]. Importantly, SARS-CoV-2 remained among the most frequently detected pathogens, supporting ongoing viral circulation and clinical relevance in the post-COVID-19 era.

Mixed infections accounted for a considerable proportion of positive cases, with *rhinovirus/enterovirus* co-detection being particularly prominent. Co-infections are increasingly recognized as a frequent finding in multiplex PCR testing, reflecting the ability of syndromic panels to identify pathogens that may be missed by conventional diagnostic workflows [15, 21]. However, interpretation of co-detections requires caution, as nucleic acid detection does not always indicate active causality and may occasionally represent prolonged shedding or incidental carriage, especially in older or hospitalized patients [17].

Finally, no statistically significant association was observed between test positivity and patient age or gender. Similar observations have been reported in other retrospective evaluations, suggesting that pathogen circulation is more strongly influenced by seasonal and epidemiological factors than by demographic variables alone within hospitalized cohorts [16]. Overall, these findings reinforce the value of Film Array-based syndromic testing as a rapid tool for pathogen surveillance and etiological clarification in routine hospital practice [8].

Importantly, negative results do not necessarily exclude infection, as syndromic panels are limited to predefined targets and may miss emerging pathogens, low viral load infections, or non-infectious causes of respiratory symptoms. Such interpretive challenges have been widely emphasized in diagnostic stewardship frameworks [19].

A major strength of our study lies in the detailed description of etiological agents circulating in the post COVID-19 period. SARS-CoV-2 remained among the most frequently detected pathogens, confirming its continued clinical impact beyond the pandemic peak. Concurrently, influenza viruses, RSV, human metapneumovirus, parainfluenza viruses, adenovirus, and seasonal corona viruses were also identified, demonstrating the re-establishment of the broader respiratory viral ecosystem. This pattern is consistent with international surveillance studies showing the resurgence of influenza and RSV following the relaxation of pandemic-related public health measures [21].

Notably, the detection of atypical bacterial pathogens such as *Mycoplasma pneumoniae* and *Bordetella pertussis*, although infrequent, highlights the diagnostic advantage of multiplex PCR assays over conventional culture-based workflows. These organisms are difficult to isolate with routine techniques, and delayed recognition may contribute to inappropriate antimicrobial exposure or missed opportunities for targeted therapy. Expert consensus documents increasingly recognize rapid multiplex syndromic testing as transformative, particularly in critically ill patients where early etiological clarification is essential [8, 15].

An important finding in our study population (patient sample) was the high proportion of mixed infections, with *rhinovirus/enterovirus* co-detection being particularly prominent. Co-infections have been repeatedly reported in international Film Array evaluations, especially among elderly and hospitalized populations, and represent a key benefit of broad multiplex testing compared with single-target PCR approaches [21]. However, co-detection also introduces interpretive complexity, since nucleic acid detection does not always imply active pathogenicity. Persistent shedding, colonization, or incidental viral carriage may complicate clinical decision making, reinforcing the need for careful correlation with symptoms and radiological findings [17].

International evidence supports that syndromic multiplex respiratory panels can provide earlier pathogen identification and higher diagnostic yield than conventional culture-based approaches, particularly in hospitalized patients [31, 32]. Rapid turnaround can facilitate earlier infection control interventions, improved cohorting of patients, and reduction of unnecessary antibiotic prescribing. Similarly, in bloodstream infections, Film Array BCID2 implementation has shown high concordance with standard methods while significantly reducing time to pathogen identification and resistance marker detection [27, 28, 33]. In stewardship-guided environments, rapid diagnostics such as the Film Array BCID2 panel have been shown to shorten time to actionable microbiological results and facilitate earlier antimicrobial optimization compared with conventional workflows [27, 28].

Although our study did not include outcome variables such as morbidity, mortality, or antimicrobial modifications, international evidence suggests that the principal value of Film Array testing lies in its ability to influence clinical decision making early in the diagnostic pathway. For example, respiratory panel testing has been linked to reductions in antibiotic days of therapy and more appropriate antiviral use in diverse clinical settings [34]. Therefore, the diagnostic patterns observed in our hospital likely carry meaningful implications for patient management even if direct outcome measures were not available.

A further contribution of this work is the demonstration of clear seasonal variation [35]. Peaks during late winter and early spring were observed, consistent with established respiratory virus circulation dynamics in temperate climates, where low temperature, indoor crowding, and humidity changes promote viral transmission. Additional increases during late summer and autumn may reflect population mixing after vacation periods and school reopening, trends also documented in international epidemiological studies [36, 37].

Continuous multiplex surveillance may therefore support hospital preparedness, staffing allocation, and infection prevention planning [38-40]. Departmental distribution of testing revealed that the majority of specimens originated from Pulmonology and Internal Medicine wards, reflecting the substantial burden of respiratory infections among hospitalized adults and elderly patients. Given the advanced mean age of the patient sample (study population), these findings align with global data demonstrating that respiratory viruses remain a major cause of hospitalization and healthcare strain in older populations even outside pandemic condi-

tions [39-42].

Clinical implications for Greece

The implementation of rapid syndromic molecular diagnostics such as Film Array has particular relevance for the Greek healthcare system [43-45].

Greece, like many Southern European countries, faces seasonal surges of respiratory infections, high antibiotic consumption rates, and increasing antimicrobial resistance pressures [46-51].

Rapid multiplex testing may support earlier differentiation between viral and bacterial etiologies, enabling improved antimicrobial stewardship and reducing unnecessary antibiotic exposure, a critical priority for national public health [52]. Moreover, hospital-based molecular surveillance can provide valuable epidemiological intelligence for regional preparedness, especially in the context of tourism-driven population mobility and climate-related shifts in respiratory virus seasonality [45, 53]. Expanding access to multiplex diagnostics across Greek hospitals could therefore strengthen both patient-level management and national infectious disease surveillance capacity [54].

Limitations

Several limitations and potential sources of bias should be acknowledged in this study. First, the retrospective single-center design restricts the generalizability of our findings and prevents causal inference regarding the clinical impact of Film Array testing. In addition, selection and referral bias may be present, as multiplex PCR was primarily requested for hospitalized patients particularly from Pulmonology and Internal Medicine wards, therefore not fully representing the broader community population or other hospital departments. Second, the absence of detailed clinical outcome data, including antimicrobial therapy modifications, length of hospitalization, morbidity, or mortality, limits the ability to evaluate patient-centered benefits or the direct influence of syndromic testing on clinical decision making. Third, multiplex PCR assays detect microbial nucleic acid rather than viable organisms; consequently, positive results may not always reflect active infection, and co-detections may occasionally represent prolonged viral shedding or incidental carriage rather than true etiological relevance. Furthermore, the restricted target range of the assay may contribute to negative results, as pathogens outside the panel remain undetectable. Finally, the uneven distribution of samples across months may have influenced the observed seasonal patterns, underscoring the importance of interpreting epidemiological trends within the context of testing practices and diagnostic stewardship [19].

Conclusions

Despite these limitations, our retrospective study provides valuable post-pandemic real-world evidence on the etiological spectrum, co-infection burden, and seasonal dynamics of respiratory pathogens detected by Film Array technology in a Greek tertiary hospital during 2024. The diversity of identified viruses, the detection of atypical bacteria, and the frequency of mixed infections underline the diagnostic utility of syndromic multiplex PCR not only for individual patient evaluation but also for hospital epidemiological surveillance. Future prospective multicenter studies incorporating antimicrobial stewardship metrics and clinical outcomes are warranted to further define the full impact of multiplex diagnostics in Greece and internationally.

Author contributions

Antonia Mourtzikou wrote the initial and final versions of the manuscript. Christina Seitopoulou conducted the laboratory analysis. Georgia Kalliora performed the literature search. Marilena Stamouli provided the statistical analysis.

All authors contributed equally to the discussion of the content and to

writing, reviewing, and editing the manuscript prior to submission.

Financial Disclosure

Financial support for the study was provided by Maria Kimouli.

Conflict of Interest

The authors declared no conflict of interest.

REFERENCES

1. Babady NE. The Film Array[®] respiratory panel: an automated, broadly multiplexed molecular test for the rapid and accurate detection of respiratory pathogens. *Expert Rev Mol Diagn.* 2013;13(8):779-88. doi: 10.1586/14737159.2013.848794. PMID: 24151847; PMCID: PMC7103684
2. Kardjadj M. Advances in Point-of-Care Infectious Disease Diagnostics: Integration of Technologies, Validation, Artificial Intelligence, and Regulatory Oversight. *Diagnostics (Basel).* 2025;15(22):2845. doi: 10.3390/diagnostics15222845. PMID: 41300871; PMCID: PMC12651474
3. Aygar İS, Hoşbul T. Diagnostic accuracy and clinical impact of Film Array multiplex PCR system in bloodstream infections: A comparative study with conventional methods in a tertiary health care setting. *Medicine (Baltimore).* 2025;104(29):e43263. doi: 10.1097/MD.00000000000043263. PMID: 40696646; PMCID: PMC12282700
4. Hassall J, Coxon C, Patel VC, Goldenberg SD, Sergaki C. Limitations of current techniques in clinical antimicrobial resistance diagnosis: examples and future prospects. *NPJ Antimicrob Resist.* 2024;2(1):16. doi: 10.1038/s44259-024-00033-8. PMID: 39843577; PMCID: PMC11721362.0
5. Edmondson R, Saeed K, Green S, O'Dwyer M. Improving Turn-around Times for Routine Antimicrobial Sensitivity Testing Following European Committee on Antimicrobial Susceptibility Testing Methodology in Patients with Bacteraemia. *Antibiotics.* 2024; 13(11):1094. <https://doi.org/10.3390/antibiotics13111094>
6. Kaprou GD, Bergšpica I, Alexa EA, Alvarez-Ordóñez A, Prieto M. Rapid Methods for Antimicrobial Resistance Diagnostics. *Antibiotics.* 2021; 10(2):209. <https://doi.org/10.3390/antibiotics10020209>
7. Pinzauti D, Biazzo M, Podrini C, Alevizou A, Safarika A, Damoraki G, et al. An NGS-assisted diagnostic workflow for culture-independent detection of bloodstream pathogens and prediction of antimicrobial resistances in sepsis. *Front. Cell. Infect. Microbiol.* 2025; 15:1656171. doi: 10.3389/fcimb.2025.1656171
8. Candel FJ, Salavert M, Cantón R, Del Pozo JL, Galán-Sánchez F, Navarro D, et al. The role of rapid multiplex molecular syndromic panels in the clinical management of infections in critically ill patients: an experts-opinion document. *Crit Care.* 2024; 28(1):440. doi: 10.1186/s13054-024-05224-3. PMID: 39736683; PMCID: PMC11687037
9. Yilmaz M, Kilic S, Bayraktar F, Ötgün SN, Tosun AI, Zeybek U, et al. Syndromic Testing-The Evaluation of Four Novel Multiplex Real-Time Polymerase Chain Reaction Panels. *Diagnostics (Basel).* 2025; 15(10):1228. doi: 10.3390/diagnostics15101228. PMID: 40428221; PMCID: PMC12109757
10. Gabrielli L, Tomaiuolo M, Banchini I, Balboni A, Liberatore A, Lanna F, et al. Performance Evaluation of Multiplex Molecular Syndromic Panel vs. Singleplex PCR for Diagnosis of Acute Central Nervous System Infections. *Microorganisms.* 2025; 13(4):892. <https://doi.org/10.3390/microorganisms13040892>
11. Sharjeel M, Irshad M, Rizvi A, Ahmed A, Leghari MJ. Multiplex PCR pneumonia panel compared to standard culture of respiratory specimens: Retrospective results from a transplant centre. *J Glob Antimicrob Resist.* 2025; 46:179-186. doi: 10.1016/j.jgar.2025.12.004. Epub ahead of print. PMID: 41371585
12. Wang L, Cai J, Dai L, Miao W, Li Z, Cao W, et al. Multicenter evaluation of fast multiplex PCR for detection of pathogens in lower respiratory tract infections. *Front. Cell. Infect. Microbiol.* 2025; 15:1643991. doi: 10.3389/fcimb.2025.1643991
13. Serapide F, Pallone R, Quirino A, Marascio N, Barreca GS, Davoli C, et al. Impact of Multiplex PCR on Diagnosis of Bacterial and Fungal Infections and Choice of Appropriate Antimicrobial Therapy. *Diagnostics (Basel).* 2025; 15(8):1044. doi: 10.3390/diagnostics15081044. PMID: 40310414; PMCID: PMC12026191
14. Gómez de la Torre Pretell JC, Hueda-Zavaleta M, Cáceres-DelAguija JA, Barletta-Carrillo C, Copaja-Corzo C, Poccorpachi MDPS, et al. Clinical Characteristics Associated with Detected Respiratory Microorganism Employing Multiplex Nested PCR in Patients with Presumptive COVID-19 but Negative Molecular Results in Lima, Peru. *Trop Med Infect Dis.* 2022; 7(11):340. doi: 10.3390/tropicalmed7110340. PMID: 36355882; PMCID: PMC9692319
15. Dumkow LE, Worden LJ, Rao SN. Syndromic diagnostic testing: a new way to approach patient care in the treatment of infectious diseases. *J Antimicrob Chemother.* 2021; 76 (3):iii4-iii11. doi: 10.1093/jac/dkab245. PMID: 34555157; PMCID: PMC8460095
16. Søgaard KK, Hinic V, Goldenberger D, Gensch A, Schweitzer M, Bättig V, et al. Evaluation of the clinical relevance of the Bio Fire[®] Film Array pneumonia panel among hospitalized patients. *Infection.* 2024; 52:173-181. <https://doi.org/10.1007/s15010-023-02080-1>
17. Riaño-Sánchez LF, Alvarez-Moreno CA, Godoy M, Sierra CR, Castañeda MI, Cortés JA. Multiplex PCR Pneumonia Panel in Critically Ill Patients Did Not Modify Mortality: A Cohort Study. *Antibiotics.* 2025; 14(3):245. <https://doi.org/10.3390/antibiotics14030245>
18. Manatrey-Lancaster JJ, Bushman AM, Caligiuri ME, Rosa R. Impact of Bio Fire Film Array respiratory panel results on antibiotic days of therapy in different clinical settings. *Antimicrobial Stewardship & Healthcare Epidemiology.* 2021;1(1):e4. doi:10.1017/ash.2021.164
19. Subedi S, Harris PNA, Hall L, Paterson DL. Practical considerations for implementation of syndromic panel and diagnostic stewardship in the era of syndromic panel testing. *Clin Microbiol Infect.* 2025; 31(11):1822-1827. doi: 10.1016/j.cmi.2025.06.013
20. Kitagawa D, Kitano T, Kasamatsu T, Shiraiishi N, Yasuda M, Okada M, et al. Impact of multiplex polymerase chain reaction testing in patients with bacteremia. *Microbiol Spectr.* 2025; 13:e01980-25. <https://doi.org/10.1128/spectrum.01980-25>
21. Hong YJ, Jung BK, Kim JK. Epidemiological Characterization of Respiratory Pathogens Using the Multiplex PCR Film Array[™] Respiratory Panel. *Diagnostics (Basel).* 2024; 14(7):734. doi: 10.3390/diagnostics14070734. PMID: 38611647; PMCID: PMC11011807
22. Babady NE, England MR, Jurcic Smith KL, He T, Wijetunge DS, Tang YW, et al. Multicenter Evaluation of the ePlex Respiratory Pathogen Panel for the Detection of Viral and Bacterial Respiratory Tract Pathogens in Nasopharyngeal Swabs. *J Clin Microbiol.* 2018; 56(2):e01658-17. doi: 10.1128/JCM.01658-17. PMID: 29212701; PMCID: PMC5786739
23. Esposito S, Mencacci A, Cenci E, Camilloni B, Silvestri E, Principi N. Multiplex Platforms for the Identification of Respiratory Pathogens: Are They Useful in Pediatric Clinical Practice? *Front Cell Infect Microbiol.* 2019; 9:196. doi: 10.3389/fcimb.2019.00196. PMID: 31275863; PMCID: PMC6593267
24. Weidmann MD, Green DA, Berry GJ, Wu F. Assessing respiratory viral exclusion and affinity interactions through co-infection incidence in a pediatric population during the 2022 resurgence of influenza and RSV. *Front Cell Infect Microbiol.* 2023; 13:1208235. doi: 10.3389/fcimb.2023.1208235. PMID: 37389220; PMCID: PMC10302716
25. López N, Cuesta G, Rodríguez-Vega S, Rosas E, Chumbita M, Casals-Pascual C, et al. Multiplex real-time PCR Film Array performance in the diagnosis of meningoencephalitis: lights and shadows. *Infection.* 2024; 52(1):165-172. doi: 10.1007/s15010-023-02076-x. Epub 2023 Jul 29. PMID: 37515691; PMCID: PMC10810907
26. Perera M, Varadhan H, Oon A. A post-implementation evaluation of Bio Fire Film Array Meningitis/Encephalitis panel for pathogen detec-

- tion in cerebrospinal fluid with a special focus on clinical significance of HHV-6. *Microbiol Spectr.* 2026; 14(1):e0062025. doi: 10.1128/spectrum.00620-25. PMID: 41313027; PMCID: PMC12772252
27. Caméléna F, Péan de Ponfilly G, Pailhoriès H, Bonzon L, Alanio A, Poncin T, et al. Multicenter Evaluation of the Film Array Blood Culture Identification 2 Panel for Pathogen Detection in Bloodstream Infections. *Microbiol Spectr.* 2023; 11(1):e0254722. doi: 10.1128/spectrum.02547-22. PMID: 36519852; PMCID: PMC9927563
 28. Peri AM, Ling W, Furuya-Kanamori L, Harris PNA, Paterson DL. Performance of Bio Fire Blood Culture Identification 2 Panel (BCID2) for the detection of bloodstream pathogens and their associated resistance markers: a systematic review and meta-analysis of diagnostic test accuracy studies. *BMC Infectious Diseases.* 2022; 22(1):794. DOI: 10.1186/s12879-022-07772-x. PMID: 36266641; PMCID: PMC9585790
 29. Tseng HY, Chen CL, Chen WC, Kuo YC, Liang SJ, Tu CY, et al. Reduced mortality with antimicrobial stewardship guided by Bio Fire Film Array Blood Culture Identification 2 panel in critically ill patients with bloodstream infection: A retrospective propensity score-matched study. *Int J Antimicrob Agents.* 2024; 64(4):107300. doi: 10.1016/j.ijantimicag.2024.107300. PMID: 39173938
 30. Edin A, Eilers H, Allard A. Evaluation of the Bio Fire Film Array Pneumonia panel plus for lower respiratory tract infections. *Infect Dis (Lond).* 2020;52(7):479-488. doi: 10.1080/23744235.2020.1755053. PMID: 32319831
 31. Clark TW, Lindsley K, Wigmosta TB, Bhagat A, Hemmert RB, Uyei J, et al. Rapid multiplex PCR for respiratory viruses reduces time to result and improves clinical care: Results of a systematic review and meta-analysis. *J Infect.* 2023; 86(5):462-475. doi: 10.1016/j.jinf.2023.03.005. PMID: 36906153
 32. Serigstad S, Markussen D, Grewal HMS, Ebbesen M, Kommedal Ø, Heggelund L, et al. Rapid syndromic PCR testing in patients with respiratory tract infections reduces time to results and improves microbial yield. *Sci Rep.* 2022; 12(1):326. doi: 10.1038/s41598-021-03741-7. PMID: 35013351; PMCID: PMC8748978
 33. Peri AM, Chatfield MD, Ling W, Furuya-Kanamori L, Harris PNA, Paterson DL. Rapid Diagnostic Tests and Antimicrobial Stewardship Programs for the Management of Bloodstream Infection: What Is Their Relative Contribution to Improving Clinical Outcomes? A Systematic Review and Network Meta-analysis. *Clinical Infectious Diseases.* 2024; 79 (2): 502–515. <https://doi.org/10.1093/cid/ciae234>
 34. Manatrey-Lancaster JJ, Bushman AM, Caligiuri ME, Rosa R. Impact of Bio Fire Film Array respiratory panel results on antibiotic days of therapy in different clinical settings. *Antimicrob Steward Healthc Epidemiol.* 2021; 1(1):e4. doi: 10.1017/ash.2021.164. PMID: 36168499; PMCID: PMC9495546
 35. Masoorian E, Teimoori A, Bakhtiari S, Jalilian FA, Vosough RN, Ansari N. Post-COVID-19 Seasonality of Influenza, Respiratory Syncytial Virus, and SARS-CoV-2 Among Hospitalized Children in Western Iran: A Molecular Surveillance Study (2023-2024). *J Epidemiol Glob Health.* 2025; 15(1):146. doi: 10.1007/s44197-025-00497-5. PMID: 41366524; PMCID: PMC12696241
 36. Liu Y, Chen Z, Zhang M, Wang D, Ma M, Qin P, et al. Vacation and back-to school effect on influenza transmission among school age children in Guangzhou, China: an ecological study from 2010 to 2023. *BMJ Open* 2025; 15:e096341. doi:10.1136/bmjopen-2024-096341
 37. Qiao J, Nishiura H. Public holidays increased the transmission of COVID-19 in Japan, 2020-2021: a mathematical modelling study. *Epidemiology and Health.* 2024; 46, Article ID: e2024025. <https://doi.org/10.4178/epih.e2024025>
 38. Al-Tawfiq JA, Zumla A, Gautret P, Gray GC, Hui DS, Al-Rabeeh AA, Memish ZA. Surveillance for emerging respiratory viruses. *Lancet Infect Dis.* 2014; 14(10):992-1000. doi: 10.1016/S1473-3099(14)70840-0. PMID: 25189347; PMCID: PMC7106459
 39. Chauvel C, Casalegno JS, Visseaux B, Vieillefond V, Haim-Boukozba S, Enouf V, et al. Community and Hospital-Based Laboratory Surveillance for Influenza, Respiratory Syncytial Virus, and SARS-CoV-2 During the 2023-2024 Season, Lyon, France. *J Med Virol.* 2025; 97(9):e70549. doi: 10.1002/jmv.70549. PMID: 40880188; PMCID: PMC12396161
 40. Ghosal K, Adhikari A. Aerobiology of Respiratory Infectious Viruses: Recent Paradoxes, Mechanistic Insights, and Future Perspectives. *Aerobiology.* 2025; 3(3):7. <https://doi.org/10.3390/aerobiology3030007>
 41. Torres AR, Gómez V, Kislaya I, Rodrigues AP, Fernandes Tavares M, Pereira AC, et al. Monitoring COVID-19 and Influenza: The Added Value of a Severe Acute Respiratory Infection Surveillance System in Portugal. *Can J Infect Dis Med Microbiol.* 2023; 2023:6590011. doi: 10.1155/2023/6590011. PMID: 36846348; PMCID: PMC9950323
 42. Huang W-H, Ho Y-F, Yeh J-Y, Liu P-Y, Huang P-H. Hospital Influenza Outbreak Management in the Post-COVID Era: A Narrative Review of Evolving Practices and Feasibility Considerations. *Healthcare.* 2026; 14(1):50. <https://doi.org/10.3390/healthcare14010050>
 43. Messacar K, Parker SK, Todd JK, Dominguez SR. Implementation of Rapid Molecular Infectious Disease Diagnostics: the Role of Diagnostic and Antimicrobial Stewardship. *J Clin Microbiol.* 2017; 55(3):715-723. doi: 10.1128/JCM.02264-16. Epub 2016 Dec 28. PMID: 28031432; PMCID: PMC5328439
 44. Kyriazopoulou E, Karageorgos A, Liaskou-Antoniou L, Koufargyris P, Safarika A, Damoraki G, et al. Bio Fire® Film Array® Pneumonia Panel for Severe Lower Respiratory Tract Infections: Subgroup Analysis of a Randomized Clinical Trial. *Infect Dis Ther.* 2021; 10:1437–1449. <https://doi.org/10.1007/s40121-021-00459-x>
 45. Comini S, Priori AM, Coppari F, Sabbatini M, Bruno C, Boattini M, et al. Integrating Syndromic Molecular Assays into Routine Diagnostic Microbiology: Benefits and Challenges. *Antibiotics.* 2026; 15(2):182. <https://doi.org/10.3390/antibiotics15020182>
 46. Mylona E, Kostourou S, Giankoula D, Spyraou E, Michopanou N, Kolokotroni C, et al. Trends in Antimicrobial Resistance at a Greek Tertiary Hospital over a 7-Year Period, Including the COVID-19 Pandemic. *Antibiotics (Basel).* 2025; 14(11):1067. doi: 10.3390/antibiotics14111067. PMID: 41301562; PMCID: PMC12649202
 47. Simonsen GS. Antimicrobial resistance surveillance in Europe and beyond. *Euro Surveill.* 2018; 23(42):1800560. doi: 10.2807/1560-7917.ES.2018.23.42.1800560. PMID: 30352641; PMCID: PMC6199866
 48. Tsioutis C., Kritsotakis E.I., Karageorgos S.A., Stratakou S., Gikas A. Trends in antimicrobial resistance and antibiotic consumption in a tertiary care hospital in Crete, Greece. *Infect. Drug Resist.* 2021;14:1501–1511
 49. European Centre for Disease Prevention and Control. (2024). ECDC country visit to Greece to discuss antimicrobial resistance issues : 15–19 April 2024. European Centre for Disease Prevention and Control. <https://data.europa.eu/doi/10.2900/1791648>
 50. Tsalidou M., Stergiopoulou T., Bostanitis I., Nikaki C., Skoumpa K., Koutsoukou T., et al. Surveillance of antimicrobial resistance and multidrug resistance prevalence of clinical isolates in a general hospital in northern Greece. *Antibiotics.* 2023; 12:1595. doi: 10.3390/antibiotics1211159
 51. GBD 2021 Antimicrobial Resistance Collaborators. Global burden of bacterial antimicrobial resistance 1990-2021: a systematic analysis with forecasts to 2050. *Lancet.* 2024; 404(10459):1199-1226. doi: 10.1016/S0140-6736(24)01867-1. PMID: 39299261; PMCID: PMC11718157
 52. Shengchen D, Gu X, Fan G, Sun R, Wang Y, Yu D, et al. Evaluation of a molecular point-of-care testing for viral and atypical pathogens on intravenous antibiotic duration in hospitalized adults with lower respiratory tract infection: a randomized clinical trial. *Clin Microbiol Infect.* 2019; 25(11):1415-1421. doi: 10.1016/j.cmi.2019.06.012.

PMID: 31229593; PMCID: PMC7173318

53. Pae R, Millest A, Tirion A, Dryden M, Lee JE, Wight N, et al. Impact of implementation of rapid syndromic molecular diagnostics on self-reported clinical and public health practice: a qualitative study in small island health services. medRxiv preprint doi: <https://doi.org/10.1101/2024.11.05.24316772>
54. Brendish NJ, Malachira AK, Armstrong L, Houghton R, Aitken S,

Nyimbili E, et al. Routine molecular point-of-care testing for respiratory viruses in adults presenting to hospital with acute respiratory illness (ResPOC): a pragmatic, open-label, randomised controlled trial. *Lancet Respir Med.* 2017; 5(5):401-11. doi: 10.1016/S2213-2600(17)30120-0

Cite this article: * Mourtzikou A, Seitopoulou C, Stamouli M, Kalliora G, Kimouli M. (2026) Detection of viral and bacterial infections by means of Film Array at a Tertiary Hospital in Greece. *Advance Medical & Clinical Research.* 7 (1): 284-290.

Copyright: ©2026 Antonia Mourtzikou. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.